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Residues Analysis of Acetamiprid, Boscalid, Imidacloprid and Pyraclostrobin in the Minor Crop Mustard Green under Greenhouse Conditions for Evaluation of their Potentiality of PLS Violation

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Abstract

BACKGROUND: The demand for pesticide registration has kept increasing for minor crop cultivation in greenhouse since Positive List System (PLS) has been launched. Thus, much study on the evaluation of pesticide residues in minor crops is required to examine the demand. In this study, we evaluated residues of acetamiprid, boscalid, imidacloprid and pyraclostrobin in the minor crop mustard green to provide the potential data for their registration.

METHODS AND RESULTS: Pesticide granule formulations of acetamiprid, boscalid, imidacloprid and pyraclostrobin were incorporated into soil and applied onto field soil surface at rates of 3 kg/10a, 6 kg/10a, 3 kg/10a and 6 kg/10a, respectively. The pesticides were also applied at the two times higher than the rates to compare the residues between the application rates. Mustard green seeds were sown 1 day after pesticide application and cultivated under greenhouse conditions. LC/MS/MS analyses coupled with a modified QuEChERS method were employed for determination of the pesticides in plant samples. The method limits of quantitation (LOQ) of the pesticides were 0.01 mg/kg, and

the matrix calibration curves of the pesticides showed linearity with coefficient values of determination (r^2) greater than 0.995. The average recovery values of the pesticides fortified in control samples at rates of LOQ and 10LOQ ranged from approximately 77.5% to 101.2% with relative standard deviation values lower than 14%. The pesticides in the mustard green samples cultivated for 53 days after sown were determined to be lower than the LOQ level.

CONCLUSION: Acetamiprid, boscalid, imidacloprid and pyraclostrobin were found at a level lower than 0.01 mg/kg in the minor crop mustard green. Thus, their residues in mustard green would not violate PLS under greenhouse conditions.

Key words: Acetamiprid, Boscalid, Imidacloprid, PLS, Pyraclostrobin

Introduction

Korea has established Positive List System (PLS) as a pesticide safety law for agricultural products in December 31, 2016 and has applied the system to crops including seed and nut products and tropical fruits, and now has enforced PLS to whole agricultural products since January 1, 2019. The number of anxious issues have arose from agricultural farmers who would receive financial

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penalty based on the violation of PLS due to unpredictable pesticide contamination of crops. The rotational crop system is a typical concern on the unpredictable pesticide contamination because the secondary crop would be contaminated with the pesticide that had been used for the first crop. In Korea, the rotational crop system has been largely performed for minor crops in greenhouses. Thus, much effort is required to allow for minor crops not to violate PLS.

Minor crops have been typically accepted as one of income crops for farmers because the crops could be harvested in a few month after seeding or transplanting. In Korea, minor crops named agricultural plants grown in overall cultivation area smaller than 1,000ha. According to Rural Development Administration (RDA) of Korea, the cultivation variety of minor crops has increased yearly, particularly in greenhouse system [1]. The need of pesticide registration for the control of plant pathogens and pest insects in minor crops are triggered by agricultural farmers in association with the variety. Thus, methods for the determination of pesticide residues in a program of pesticide registration are essentially required.

Determination of pesticides in crops is of important not only for protection of human health but also for regulatory control of agricultural foods. QuEChERS method has been currently introduced as a powerful tool for sample preparation in a program of pesticide residue analysis [2, 3]. The method is based on reducing organic solvent use and sample preparation steps such as extraction, clean-up and concentration. QuEChERS methods have been well coordinated to mass spectrometry as multiresidue analytical methods for the determination of pesticides in agricultural food stuffs [4-6]. Acetamiprid, boscalid, imidacloprid and pyraclostrobin are one of pesticides widely used to control pest insects and plant pathogens in greenhouse [7-11]. According to National Agricultural Products Quality Management Service (NAQS), they are one of the major pesticides that have been detected in agricultural soil samples by yearly domestic pesticide monitoring program. NAQS has particularly interested in imidacloprid and acetamiprid because they were found in rotational minor crop such as perilla leaf. Giving the situation, acetamiprid, boscalid, imidacloprid and pyraclostrobin are now in a situation that Korean government should consider the registration for their use in rotational minor crops because agricultural food safety becomes an important issue due to their potentiality of unpredictable pesticide contamination

against PLS law. Thus, much effort is required to examine these pesticide residues in rotational minor crop. In this study, we used modified QuEChERS methods coupled with LC/MS/MS to examine the residues of acetamiprid, boscalid, a pyraclostrobin metabolite BF 500-3, imidacloprid and pyraclostrobin in mustard green samples cultivated under greenhouse conditions. Based on the experimental data, we evaluated potential violation of the pesticides against PLS.

Materials and Methods

Chemicals and Reagents

Acetamiprid (purity 100%), boscalid (purity 99.5%), imidacloprid (purity 100%), pyraclostrobin (purity 99.9%), and a pyraclostrobin metabolite BF500-3 (purity 99.9%) were purchased from Sigma-Aldrich Corporation (St Louis, MO, USA). Their chemical structures are presented in Fig. 1. All solvents were HPLC grade purchased from J.T. Baker Chemicals (Phillipsburg, NJ, USA). QuEChERS kits and SPE cartridge columns were obtained from Agilent Technologies (CA, USA). All chemicals were reagent grade purchased from Fisher Chemical (PA, USA) and Junsei Chemical Co. Ltd (Tokyo, Japan), unless otherwise stated.

Field Trials

Field trials were performed at a greenhouse (Damyang, Jeonnam) to investigate the residues of acetamiprid, boscalid, imidacloprid, pyraclostrobin and BF500-3 in the mustard green samples. The trials were conducted in the soil treated with the pesticides at a level highly exposable to the secondary crop. Pesticide granule formulation (GR) was chosen as a test formulation type because it was hypothesized to find the residue level at a level higher than other types in the soil after used in the first crop. Commercial GRs were mixed thoroughly with a small portion of soil at a ratio of 1 to 30 (g/g) in polyvinyl bags and treated evenly onto the greenhouse soil (2 m x 5 m) at the recommended dose levels of 6 kg/10a of Bellisplus[®] (Kyung Nong Co., Korea) for boscalid (a.i. 0.8%) and pyraclostrobin (a.i. 0.25%) and 3 kg/10a of Veteran[®] (Dongbang Agro Co., Korea) for imidacloprid (a.i. 2.0%) and 3 kg/10a of Mospiran[®] (Kyung Nong Co., Korea) for acetamiprid (a.i. 1.5%), respectively. The treated soils were then carefully mixed by using a Dongyang Techtool (Daegu, Korea) model KM WG420 agricultural farm management machine. In separate experiments, each pesticide was treated at rates two times higher than the recommended dose level. Commercial mustard green seeds

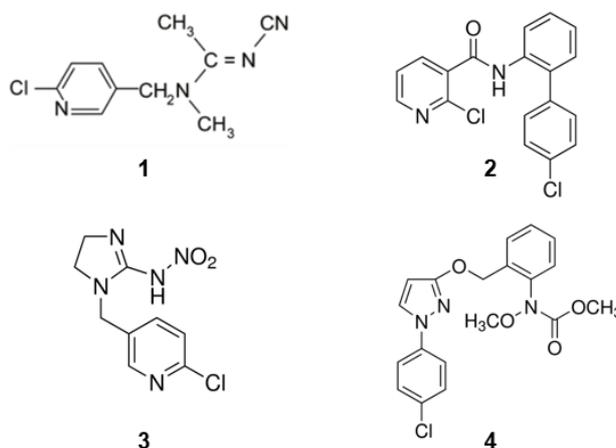


Fig. 1. Chemical structures of acetamiprid (1), boscalid (2), imidacloprid (3), and pyraclostrobin (4) examined in this study.

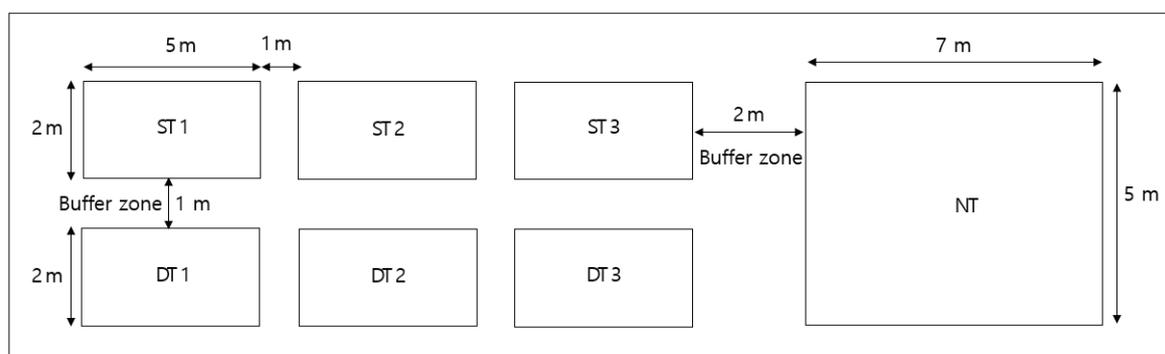


Fig. 2. Experimental design of the plots for the greenhouse experiments. ST: single dose treatment, DT: double dose treatment, NT: not treated (control).

(red leaf mustard) purchased from Nongwoo Bio (Suwon, Korea) were sown 1 day at a distance of 25 cm between plants after the pesticide treatment according to the manufacturer's recommendation. The experimental plots for the greenhouse experiments were designed in triplicate as presented in Fig. 2.

Mustard greens were cultivated conventionally for 53 days under greenhouse conditions. Mustard green samples were collected from the field trial greenhouse and then chopped into small pieces followed by grinding fully using a high-speed homogenizer. Soil samples were collected by using a model SL09018 stainless soil auger (Shinill Science INC., Korea) at a depth of 0-20 cm from eight points in each plot after the harvest. The soil samples were then air-dried overnight and passed through a 2-mm sieve. All samples were then stored at -20°C until used.

Sample Preparation

Sample preparation for pesticide analyses in plant and

soil samples were performed with methods modified from QuEChERS, as described below. The methods were optimized by modifying several factors such as extraction solvent, extraction time, centrifugation and extraction kit.

For acetamiprid, a 10 g-sample was added with 10 mL of acetonitrile to a 50 mL centrifuge tube. The sample was vortexed vigorously for 2 min and mixed with 6.0 g of anhydrous MgSO_4 and 1.5 g of NaOAC followed by vortexing again for 2 min. The mixture was centrifuged at 4,500 rpm for 5 min to obtain supernatant. A 1 mL-aliquot of supernatant was transferred into a 2 mL micro-centrifuge tube containing anhydrous MgSO_4 (150 mg), PSA (25 mg) and GCB (2.5 mg), mixed for 2 min and centrifuged at 10,000 rpm for 5 min. The resultant supernatant was used for LC/MS/MS analysis after being passed through a $0.2\ \mu\text{m}$ syringe membrane filter (PTFE-H).

For boscalid, a 10 g-sample was added with 10 mL of acetonitrile to a 50 mL centrifuge tube and vortexed vigorously with 6.0 g of anhydrous MgSO_4 and 1.5 g of

NaOAC for 2 min. The mixture was then vortexed for 2 min and centrifuged at 4,500 rpm for 5 min. An aliquot (1.0 mL) of supernatant was transferred into a 2 mL micro-centrifuge tube and mixed with anhydrous MgSO₄ (150 mg), PSA (25 mg) and GCB (2.5 mg) for 2.0 min. The mixture was centrifuged at 10,000 rpm for 5 min, and the resultant supernatant was used for LC/MS/MS analysis as described above.

For pyraclostrobin, a 10 g-sample was added to a 50 mL centrifuge tube. The sample was added with 10 mL of acetonitrile and mixed vigorously for 2 min together with 4.0 g of anhydrous MgSO₄, 1.0 g of NaCl, 1.0 g of trisodium citrate dehydrate and 0.5 g of disodium hydrogen citrate sesquihydrate. The mixture was vortexed for 2 min and centrifuged at 4,500 rpm for 5 min. A 1.0 mL-aliquot of supernatant was transferred into a 2 mL micro-centrifuge tube and mixed with anhydrous MgSO₄ (150 mg), PSA (25 mg) and GCB (2.5 mg) were added to the tube. The mixture was then vortexed for 2 min and centrifuged at 10,000 rpm for 5 min. The resultant supernatant was used for LC/MS/MS analysis as described above. Sample preparation for BF500-3, a pyraclostrobin metabolite, was performed as described for pyraclostrobin, except not using GCB.

Sample preparation for imidacloprid was conducted by extracting sample (10 g) with 10 mL of acetonitrile containing 4.0 g of anhydrous MgSO₄, 1.0 g of NaCl, 1.0 g of trisodium citrate dehydrate and 0.5 g of disodium hydrogen citrate sesquihydrate in a 50 mL centrifuge tube for 2 min. The extracts was centrifuged at 4,500 rpm for 5 min, and a 1.5 mL-aliquot of supernatant was transferred into a 2 mL micro-centrifuge tube containing anhydrous MgSO₄ (150 mg), PSA (25 mg) and GCB (7.5 mg). The mixture was vortexed for 2 min and centrifuged at 10,000 rpm for 5 min. The resultant supernatant was used for LC/MS/MS analysis.

Chemical stability of the pesticides in the samples during storage at -20°C was examined by calculating the recovery data of the standard solutions spiked in the

control samples at a level of 0.1 mg/L. The spiked samples were stored at -20°C during the same period as the field sample storage period until analyzed.

Method Validation

Method validation was performed based on ion ratio, linearity, sensitivity, precision, and accuracy, following the guidance of European Commission document SANTE/11813/2017. Default maximum permitted tolerances of the ion ratio was set as within $\pm 30\%$ of relative average of calibration standards from the sequence analysis. A qualitative ion ratio was employed to indicate the presence of interferences, and it was calculated as the ratio of the peak area for the quantifier transition to the peak area of qualifier transition in calibration standard and sample matrix solutions. The selected ions used in calculating the ion ratio are presented in Table 1. The linearity of standard solutions of the pesticides was obtained from matrix-matched calibration curve by diluting serially their stock solutions (100 mg/L) with the extracts of control samples into their working solutions ranged from 2.5 $\mu\text{g/L}$ to 200 $\mu\text{g/L}$. The method sensitivity was determined by the quantification of limit (LOQ) at the signal to noise of 10:1. The LOQ was calculated as: $\text{LOQ (mg/kg)} = [\text{minimum detectable amount (ng)} \times \text{final sample volume (mL)}] / [\text{injection volume (}\mu\text{L)} \times \text{sample amount (g)}]$. The precision and accuracy of the method were assessed by calculating the recovery data using the standard solutions fortified in the control samples at levels of LOQ and 10LOQ. Recovery data were accepted when the values were in the range of 70 - 120% with the relative standard deviation (RSD) less than 20%. Recovery was performed in triplicate, and the data were calculated as: $\text{recovery (\%)} = (\text{the concentration detected in sample}) / (\text{the fortified concentration}) \times 100$. Particularly, the concentrations of pyraclostrobin in the samples were calculated as following,

Table 1. Instrument parameters for analysis of pesticides in multiple reaction mode

Pesticide	Precursor ions (m/z)	Product ion (m/z) and collision energy (eV)	
		Quantitation	Qualitation
Acetamiprid	223.1	125.9 (21)	56.0 (23)
Boscalid	343.0	306.9 (22)	139.9 (22)
Imidacloprid	256.2	209.1 (15)	175.1 (23)
Pyraclostrobin	388.1	194.2 (20)	162.8 (11)
BF500-3	358.1	326.0 (16)	164.0 (17)

$$P = P + (B \times F)$$

P: residue concentration of pyraclostrobin (mg/kg)

B: residue concentration of BF500-3 (mg/kg)

F: conversion factor, molecular weight (MW) of pyraclostrobin / MW of BF500-3 = 387.83 / 357.80 = 1.08

Instrument

LC/MS/MS was a Waters model Xevo TQ-XS triple quadrupole mass spectrometer equipped with a Waters model ACQUITY™ UPLC system. An Osaka Soda CAPCELL CORE C18 stainless column (150 x 2.1 mm, 2.7 μm thickness) was used for the chromatographic separation. The mobile phase was consisted of acetonitrile (A) and water (B) containing 0.1% (v/v) formic acid, and its isocratic and gradient were performed as follows: for acetamiprid, isocratic 15% solvent A for 1.0 min, linear gradients 60% solvent A for 1.5 min and 90% solvent A for 10 min, isocratic 90% solvent A for 10 min, flow rate 0.3 mL/min; for boscalid, isocratic 60% solvent A, flow rate 0.2 mL/min; for imidacloprid, isocratic 5% solvent A for 0.25 min, linear gradient 100% solvent A for 7.75 min, isocratic 100% solvent A for 8.5 min, flow rate 0.45 mL/min; for BF500-3 and pyraclostrobin, isocratic 50% solvent A for 3.0 min, linear gradient 80% solvent A for 9.0 min, isocratic 80% solvent A for 13 min, flow rate 0.3 mL/min. LC/MS/MS spectra were acquired with the positive ion in electron spray ionization (ESI) mode. The routine conditions of LC/MS/MS were optimized and set as de-solvation gas flow 10.8 mL/min (N₂), capillary gas

flow 0.34 mL/min, capillary voltage 3 KV, ion source temperature 150°C and de-solvation temperature 350°C.

Results and Discussion

The methods for sample preparation and LC/MS/MS parameters were validated in order to evaluate the residues of pesticides in mustard green samples obtained from field trails. The validation was examined based on the linearity, ion ratio, sensitivity, precision and accuracy, following the guidance of European Commission document SANTE/11813/2017. Table 2 shows the linear equations, coefficient values of determination (r^2), ion ratios, and matrix effects of standard solutions. All pesticides showed r^2 values greater than 0.995 in matrix standard solution. The matrix effects were obtained from the slopes of linear relationships in between solvent only and matrix-matched calibrations, as matrix effect (%) = (slope of linearity curve in matrix - slope of linearity curve in solvent only) / slope of linearity curve in solvent only × 100% [12]. The matrix effects ranged from average -61.642 to 24.719%, suggesting that the matrix effect should be considered for standard calibration. If the matrix effect is lower than ±10%, the matrix can be ignored [13]. Thus, the matrix-matched calibration was employed in this study because the effects were higher than ±10%. Ion ratios of the peak area for the quantifier transition to the peak area of qualifier transition for matrix solutions ranged from approximately 0.13 to 2.31, demonstrating the

Table 2. Linear equations, coefficient values of determinations (r^2), ion ratios and matrix effects of pesticide standard solutions

Matrix	Pesticide	Linear slope equation	r^2	Ion ratio ¹⁾	Matrix effect (%) ²⁾	Ion ratio tolerance (%) ³⁾	LOQ (mg/kg)
Neat solvent	Acetamiprid	$y = 20.493x + 36.711$	0.9993	0.206	-	-	-
	Boscalid	$y = 11.956x - 12.189$	0.9953	0.405	-	-	-
	Imidacloprid	$y = 13.083x + 25.737$	0.9973	0.822	-	-	-
	Pyraclostrobin	$y = 70.879x + 80.044$	0.9973	2.381	-	-	-
Mustard green	BF500-3	$y = 143.29x + 283.71$	0.9994	0.128	-	-	-
	Acetamiprid	$y = 7.6557x + 5.7285$	0.9994	0.242	-62.642	17.476	0.01
	Boscalid	$y = 12.504x - 7.8278$	0.9990	0.400	-2.358	-1.234	0.01
	Imidacloprid	$y = 14.097x + 17.731$	0.9998	0.806	7.751	-1.946	0.01
	Pyraclostrobin	$y = 73.869x + 33.34$	0.9999	2.314	4.218	-2.814	0.01
	BF500-3	$y=178.71x+496.72$	0.9995	0.129	24.719	0.781	0.01

¹⁾ Peak area of qualification ion/peak area of quantitation ion.

²⁾ [(Slope of linearity curve in matrix - slope of linearity curve in solvent only)/(slope of linearity curve in solvent only)] × 100%.

³⁾ (Average ion ratio in sample - average ion ratio in solvent)/(average ion ratio in solvent) × 100.

presence of target pesticides in the samples as shown in Fig. 3. Pyraclostrobin showed the highest ion ratio among the pesticides. In this study, the default maximum permitted tolerances of the ion ratio was set as within $\pm 30\%$ during the sequence analysis, following to the guidance of the document SANTE/11813/2017. The ion ratio tolerance ranged from -2.895 to 17.476% depending on the pesticide (Table 2), finding the ratios within $\pm 30\%$. These results reveals that the presence of the pesticides was confirmed really in the sample solutions.

Table 3 shows the data for the recovery values of the pesticides obtained from the fortified samples at LOQ and 10LOQ levels. The LOQ level was 0.01 mg/kg for all pesticides at the signal to noise of 10:1. The recovery values ranged from average 77.5 to 98.4% at LOQ level with relative standard deviation (RSD) values lower than approximately 14% while the values ranged from average 79.2 to 101.2% at 10LOQ level with RSD values lower than about 5%. These precision was acceptable accuracy

to the guidance of document SANTE/11813/2017. Overall, the results indicated that the methods examined in this study were validated sufficiently in terms of sensitivity, precision and accuracy, which meets the requirement for pesticide residue analysis in the program of Crop Residue Analysis of Minor Crop for Pesticide Registration Authority of Rural Development of Administration (RDA), Republic of Korea.

The methods validated above were apply for the determination of acetamiprid, boscalid, BF500-3, imidacloprid and pyraclostrobin in mustard green products cultivated under greenhouse conditions for 53 days after sown. Table 4 shows the concentrations of the target pesticides found in the samples. LC/MS/MS analyses detected the pesticide residues lower than the LOQ level (0.01 mg/kg) in the samples treated with the pesticides single and double dose levels. Particularly, pyraclostrobin was detected at the LOQ level even if the residue amount of its metabolite was combined

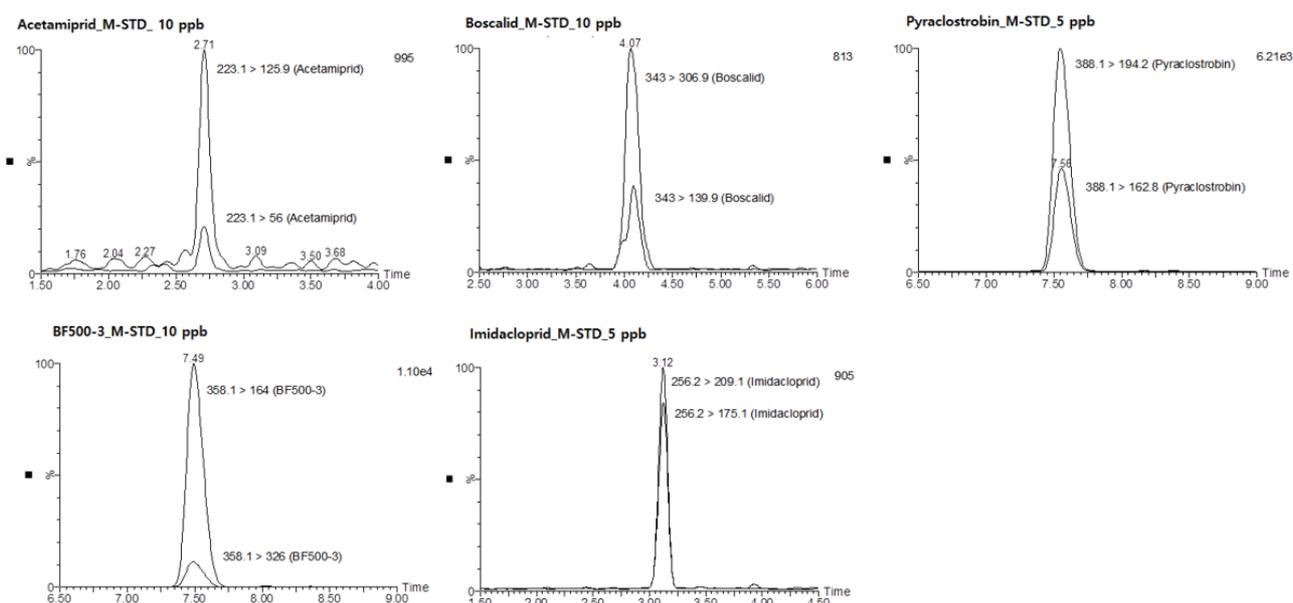


Fig. 3. Typical LC/MS/MS spectra of pesticides at minimum detectable levels examined in this study.

Table 3. Recovery values of pesticides fortified in control mustard green samples

Pesticide	Recovery values (%) [*]	
	1LOQ level (0.01 mg/kg)	10LOQ level (0.1 mg/kg)
Acetamiprid	88.1 \pm 12.3	96.0 \pm 4.3
Boscalid	77.5 \pm 1.1	92.7 \pm 4.6
Imidacloprid	98.4 \pm 5.6	101.2 \pm 3.4
Pyraclostrobin	81.0 \pm 2.4	79.2 \pm 3.3
BF500-3	80.5 \pm 4.8	96.7 \pm 1.1

* Means \pm SD of triplicate.

Table 4. Residue concentrations of pesticides in mustard green samples cultivated under greenhouse conditions

Pesticide	Concentration (mg/kg) ¹⁾	
	Single treatment	Dual treatment
Acetamiprid	<0.01	<0.01
Boscalid	<0.01	<0.01
Imidacloprid	<0.01	<0.01
Pyraclostrobin ²⁾	<0.01	<0.01

¹⁾ Means of triplicate.

²⁾ $P = P + (B \times F)$

P: residue concentration of pyraclostrobin (mg/kg)

B: residue concentration of BF500-3 (mg/kg)

F: conversion factor, molecular weight (MW) of pyraclostrobin / MW of BF500-3 = 387.83 / 357.80 = 1.08

together. The pesticides were chemically stable during the experiment, which was confirmed by storage stability tests (Supplemental S1). Supplemental experiments were performed in order to investigate pesticide residues in soil samples after plant harvest. The data showed the residue levels ranged from approximately 0.01 mg/kg to 0.9 mg/kg (Supplemental S2). Although the transport ratio from the soil to the plant was not evaluated in this study, low concentration of the pesticides in the soil would result in low residue level in the plant samples. Boscalid has been reported to degrade rapidly in 28 days after treated in soil at 3.0 mg/kg and 6.0 mg/kg, resulting in approximately 72 to 82% degradation of initial levels, in which the uptake ratio of boscalid from the soil to cabbage was lower than 2.4% of the initial level found in the soil [14]. European Food Safety Authority (EFSA) demonstrated that the residues of pyraclostrobin in rotational crops resulting from the soil uptake are not expected to be higher than 0.01 mg/kg [15]. In a field study of imidacloprid, the residue of imidacloprid was reported to be below the LOQ of 0.01 mg/kg in rotational crops after bare soil application [16]. A study of acetamiprid on rotational crops was suggested not to require because it is rapidly degraded in soil, giving the 90% degradation time (DT_{90}) of approximately 20 days [17]. These previous studies suggested that the pesticides tested in this study are generally degraded in a short time in agricultural products including plant and soil, contributing to the residue level of 0.01 mg/kg as obtained from this study. Overall, our study suggested that acetamiprid, boscalid, imidacloprid and pyraclostrobin treated at their recommended application levels would not violate PLS since their residue levels in the plant samples were lower than 0.01 mg/kg.

PLS is one of worldwide regulation guidelines against

pesticide residue violation in agricultural products. The domestic and imported products containing unregistered pesticide at a level higher than 0.01 mg/kg are not allowed to sale or distribute under PLS in Korea. A number of countries belonging to OECD have performed PLS in order to control more strongly pesticide residue in agricultural product, and Korea has applied PLS to whole agricultural products since January 1, 2019. However, PLS has generated a number anxious issues for agricultural farmers who would accidentally violate PLS due to unpredictable pesticide contamination through rotational crop, drift and groundwater. Particularly, the rotational crop system has a high potentiality of unpredictable pesticide contamination because the pesticide that had been used for the first crop would be taken and transported by the secondary crop. In case of the pesticides used for the first crop are not registered for the second crop and found at a level higher than 0.10 mg/kg, the second crop becomes to violate PLS. Thus, much effort is required to resister pesticides available for the second crops. In this study, we examined the residues of acetamiprid, boscalid, imidacloprid and pyraclostrobin in mustard green, a typical rotational minor crop, under greenhouse conditions in order to document basic data officially required for their registration. The methods for the determination of the target pesticides in the mustard green samples were optimized to the guidance of the document SANTE/11813/2017 that is required officially by RDA for pesticide residue analysis. In conclusion, based on experimental data, the target pesticides have little potential to violate PLS in mustard green.

Note

The authors declare no conflict of interest.

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